

Preliminary communication

Adriamycin analogs hydroxylated at C-3': synthesis and antitumor activity*

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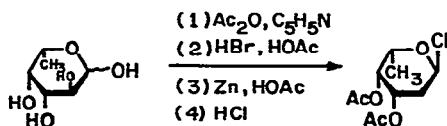
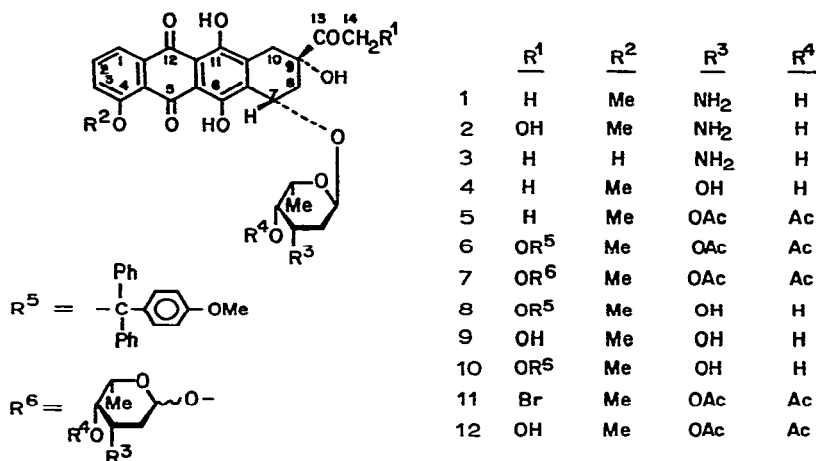
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The antitumor drugs daunorubicin (1), adriamycin (2), and carminomycin (3) have been extensively documented^{1,2}; their clinical efficacy is well established, but a cumulative, dose-related cardiotoxicity and mutagenic properties remain drawbacks in their use. In a quest for active, but less toxic analogs³, we have synthesized^{1,2} the 3'-hydroxy analog (4) of 1, and have shown² that it, and also its 3',4'-diacetate (5), demonstrate high antitumor activity, but lower toxicity than 1. As adriamycin (2) is a more potent drug than daunorubicin (1), the synthesis and evaluation of the corresponding adriamycin analogs (9 and 12), remained a significant goal, and this work is the subject of the present communication.

Three alternative approaches to 9 were envisaged: (a) from adriamycinone by sequential protection of the 14-hydroxyl group, glycosylation at O-7, and deprotection; (b) from 7-*O*-(3,4-di-*O*-acetyl-2,6-dideoxy- α -L-*lyxo*-hexopyranosyl)daunomycinone² (5) by bromination at C-14, followed by hydrolysis of the 14-bromide; and (c) by successive bromination of daunomycinone at C-14, glycosylation at O-7, and hydrolysis of the 14-bromide. It is shown that route (a) produces only mediocre yields of 9 in a tedious synthesis, and route (b) gives very poor results, whereas route (c) is effective and affords excellent yields in all steps to the biologically active diacetate 12.

For route (a), adriamycinone was obtained by acid hydrolysis⁴ of adriamycin (2) and converted into its 14-(*p*-anisylidiphenylmethyl) ether⁵. This product was treated with 2 molar equivs. of 3,4-di-*O*-acetyl-2,6-dideoxy- α -L-*lyxo*-hexopyranosyl chloride⁶ (13) under Koenigs–Knorr conditions² (yellow mercuric oxide and mercuric bromide) in the presence of powdered molecular sieve 3A in dry dichloromethane, for 24 h at 25°, to give a mixture containing two red products (t.l.c.). The excess of 13 was decomposed by addition of methanol, inorganic material was filtered off, and the products were resolved on a column of silica gel G that was eluted with 3:1 ether–petroleum ether (to remove sugar contaminants) and then with acetone to give red material, which was resolved on a second column with 4:1 benzene–acetone as the eluant. The first fractions

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afforded the desired 14-*O*-(*p*-anisyl-diphenylmethyl)-7-*O*-(3,4-di-*O*-acetyl-2,6-dideoxy- α -L-*lyxo*-hexopyranosyl)adriamycinone* (6) in 70% yield, m.p. 147–148° (chloroform–hexane), $[\alpha]_D^{22} + 115^\circ$ (c 0.01, chloroform); n.m.r. (acetone-*d*₆): δ 5.44 (bs, W_h 7 Hz, H-1' α), 7.30–7.70 (aryl), 6.92 (d, $J_{2,3}$ 9 Hz, H-3), 4.9–5.1 (m, H-3',4',7), 4.45 (broad s, 2 H, H-14), 4.2 (m, H-5'), 3.94 (s, 3 H, OCH₃), 2.74 (broad s, 2 H, H-10), 2.06, 2.09 (2 s, OAc), 1.8–2.0 (m, H-2',8), and 0.94 (d, 3 H, $J_{5',6'}$ 5 Hz, H-6'); $\nu_{\text{max}}^{\text{KBr}}$ 2.87 (OH), 5.70 (OAc, C-acetyl), 6.16, and 6.30 μm (chelated quinone).

The second fraction afforded the 7,14-bis(glycosyl)ated derivative 7 in 17% yield; m.p. 148–149° (acetone–hexane), $[\alpha]_D^{22} + 54^\circ$ (chloroform). The ¹H-n.m.r. spectrum of 7 (chloroform-*d*) indicated that the product had the sugar residue α -L-linked at O-7, but was probably a mixture of α - and β -L-substituted anomers at O-14; no signals attributable to the *p*-anisyl-diphenylmethyl group were present. Compound 7 is probably formed by 14-deprotection of either 6 or the protected adriamycinone, with subsequent glycosylation. Trial experiments with only 1 equiv. of 13 at ~25 or 0° still showed some formation of 7. No comparable 14-deprotection was reported⁵ in a synthesis of adriamycin (2) employing 2,3,6-trideoxy-4-*O*-(*p*-nitrobenzoyl)-3-(trifluoroacetamido)- α,β -L-*lyxo*-hexopyranosyl chloride. Compound 7 could be *O*-deacetylated to give 10.

O-Deacetylation of 6 was effected with cold, 0.2M sodium hydroxide, to yield

*All new compounds gave acceptable elemental analyses, and mass and 100-MHz, ¹H-n.m.r. spectra consistent with the structures reported.

35% of the partially deprotected analog 8, m.p. 152–153°, $[\alpha]_D^{22} + 80^\circ$ (chloroform); n.m.r. (CDCl_3): δ 5.44 (bs, W_h 7.5 Hz, H-1' α), 3.80 (OMe), and 7.2–7.6 (aryl).

14-Deprotection of 8 with 4:1 acetic acid–water⁵ gave “3'-hydroxyadriamycin” (9) in low yield; m/e (chemical ionization, isobutane): 337 (MH^+ – glycon – C-13,14 fragment) and 131 (glycosyl cation).

Approach (b) did not furnish a practical synthesis of 9 or 12. Treatment of compounds² 4 or 5 with bromine under a variety of conditions failed to effect a high-yielding conversion into the corresponding 14-bromo derivatives; the reactions led to complex mixtures, and the principal reaction generally involved scission of the sugar residue.

The third approach, (c), provided the most satisfactory synthesis of the target compound as its diacetate 12. 14-Bromodaunomycinone⁵ was coupled with 13 under the same conditions² used for route (a) to afford 14-bromo-7-*O*-(3,4-di-*O*-acetyl-2,6-dideoxy- α -L-*lyxo*-hexopyranosyl) daunomycinone (11) in yields of 75–94%; m.p. 152–156°, $[\alpha]_D^{25} + 119^\circ$ (chloroform); ^1H -n.m.r. (CDCl_3): δ 5.63 (d, $J_{1',2'}$ 3.1, $J_{1',2''} < 1$ Hz, H-1' α), 4.09 (s, OCH_3), 2.18, 1.95 (s, OAc), and 1.21 (d, $J_{5',6'}$ 7.1 Hz, H-6'). For consistently high yields of 11, it was necessary to use ~ 3 equiv. of 13.

Displacement of the 14-bromo group from 11 was readily achieved by the action of 5% potassium carbonate in 2:1 oxolane–water at 0°, to give a 95% yield of 7-*O*-(3,4-di-*O*-acetyl-2,6-dideoxy- α -L-*lyxo*-hexopyranosyl) adriamycinone (12), m.p. 154–157° [from acetone–petroleum ether (b.p. 30–60°)], $[\alpha]_D^{25} + 138^\circ$ (chloroform); ^1H -n.m.r. (CDCl_3): δ 5.61 (d, $J_{1',2'}$ 2.8 Hz, $J_{1',2''} < 1$ Hz, H-1' α), 13.98, 13.20 (s, phenolic OH), 4.09 (s, OCH_3), 2.18, 1.94 (s, OAc), and 1.21 (d, $J_{5',6'}$ 7.3 Hz, H-6'). *O*-Deacetylation of 12 did not afford good yields of 9 under a variety of conditions; loss

TABLE I

ANTITUMOR ACTIVITY *in vivo* OF ADRIAMYCIN AND DAUNORUBICIN ANALOGS AGAINST MURINE P-388 LYMPHOCYTIC LEUKEMIA^a

Compound	NSC Number	Dose, mg/kg ^b	T/C percent ^c
Daunorubicin (1)	82151	8	151
Adriamycin (2)	123127	4	149
3'-Hydroxydaunorubicin ² (4)	284682	200	192
3'-Hydroxydaunorubicin 3',4'-diacetate ² (5)	283158	200	186
7	311156	50, 12.5, 6.25, 3.13	100
11	307989	25	109
3'-Hydroxyadriamycin	307990	50	269
3',4'-diacetate (12)		50 ^d	202 ^d
		12.5 ^e	203 ^e
		12.5 ^f	280 ^f

^a Data obtained by A. D. Little, Inc., under the auspices of DCT, NCI. ^b CDF₁ mice were injected i.p. with 10⁶ P-388 cells on day 0, and treated i.p. on days 5, 9, and 13 with the drug dose specified. ^c Ratio (%) of median survival-time of treated mice to that of untreated controls; dose level giving maximum T/C. ^d In B-16 melanocarcinoma; single drug injection. ^e In L-1210 lymphoid leukemia; 9 daily injections. ^f In Lewis lung carcinoma.

of the sugar substituent was a major pathway under conditions² that afforded a high yield of **4** from its 3',4'-diacetate **5**.

Compound **12** shows significant, *in vivo* activity against murine P-388 lymphocytic leukemia (see Table I) and is undergoing extensive, antitumor and toxicological screening. As the 3'-hydroxyl analog (**4**) of daunorubicin and its 3',4'-diacetate (**5**) show² comparable antitumor activities, it is expected that **12** and its *O*-deacetylated analog **9** will have similar activities. Substitution for achievement of greater lipophilicity has caused enhancement⁷ of activity in the daunorubicin series, and esterases may be expected to effect the *in vivo* conversion of **12** into **9** readily.

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REFERENCES

- 1 For key references, see E.-F. Fuchs, D. Horton, and W. Weckerle, *Carbohydr. Res.*, **57** (1977) C36–C39.
- 2 E.-F. Fuchs, D. Horton, W. Weckerle, and E. Winter-Mihaly, *J. Med. Chem.*, **22** (1979) 406–411.
- 3 T. -M. Cheung, D. Horton, and W. R. Turner, *Abstr. Pap. ACS/CSJ Chem. Congr.*, Honolulu, Hawaii, April 2–5, 1979, CARB-76.
- 4 Compare, F. Arcamone, G. Cassinelli, G. Franceschi, R. Mondelli, P. Orezzi, and S. Penco, *Gazz. Chim. Ital.*, **100** (1970) 949–989.
- 5 T. H. Smith, A. N. Fujiwara, W. W. Lee, H. Y. Wu, and D. W. Henry, *J. Org. Chem.*, **42** (1977) 3653–3660.
- 6 H. S. El Khadem, D. L. Swartz, J. K. Nelson, and L. A. Berry, *Carbohydr. Res.*, **58** (1977) 230–234, and references cited therein.
- 7 G. L. Tong, H. Y. Wu, T. H. Smith, and D. W. Henry, *J. Med. Chem.*, **22** (1979) 912–918.